



Europäisches Patentamt
European Patent Office
Office européen des brevets

Publication number:

0 308 147
A1

EUROPEAN PATENT APPLICATION

Application number: 88308370.1

Int. Cl. 4: A61K 39/36 , C07K ~~45/40~~ ^{14/415}
A61K 47/00

Date of filing: 09.09.88

Priority: 12.09.87 JP 228781/87
26.07.88 JP 184487/88

Date of publication of application:
22.03.89 Bulletin 89/12

Designated Contracting States:
DE FR GB

Applicant: KABUSHIKI KAISHA HAYASHIBARA
SEIBUTSU KAGAKU KENKYUJO
2-3, 1-chome, Shimoishii
Okayama-shi Okayama(JP)

Inventor: Matsuhashi, Tyoku
6-4, 2-chome Saiwaicho
Kawaguchi-shi Saitama(JP)
Inventor: Usui, Mitsuko
7-10-102, 1-chome Okuda
Okayama-shi Okayama(JP)
Inventor: Mitsuhashi, Masakazu
4-11, 1-chome Kobashicho
Okayama-shi Okayama(JP)
Inventor: Ando, Shunsaku
291-6, Higashi-hanajiri
Okayama-shi Okayama(JP)

Representative: Pendlebury, Anthony et al
Page, White & Farrer 5 Plough Place New
Fetter Lane
London EC4A 1HY(GB)

Hyposensitization agent.

A novel hyposensitization agent was prepared by covalently attaching a saccharide, e.g. homo- and heteroglycans, for example, starch, amylose, dextran, polysucrose, pullulan, elsinan, curdlan, gum arabic, gum tragacanth, guar gum, xanthan gum, carrageenan, pectin, cellulose, glucomannan, chitosan, and lipopolysaccharide, and their derivatives and partial hydrolysates to, a cedar pollen allergen. The hyposensitization agent can be administered to a cedar pollinosis patient without fear of eliciting anaphylaxis and allergy within a shortened hyposensitization period because the hyposensitization agent much more enhances the producibilities of immunoglobulin G and M antibodies which are specific to intact cedar pollen allergen, but extremely reduces the producibility of immunoglobulin E antibody which is specific to the allergen and responsible for anaphylaxis and allergy.

EP 0 308 147 A1

HYPOSENSITIZATION AGENT

Background of the Invention1. Field of the invention

The present invention relates to a hyposensitization agent. More particularly, the present invention relates to a hyposensitization agent comprising a cedar pollen allergen covalently attached to a saccharide.

2 Abbreviations

Ala: alanine residue

Arg: arginine residue

Asn: asparagine residue

Asp: aspartic acid residue

Gln: glutamine residue

Gly: glycine residue

Ile: isoleucine residue

Lys: lysine residue

Met: methionine residue

Pro: proline residue

Ser: serine residue

Trp: tryptophan residue

Each amino acid residue is L-configuration.

3 Description of the prior art

Cedar pollinosis is an allergic disease caused by a cedar pollen scattered from blooming cedars.

Recently, the number of cedar pollinosis patients is gradually increasing in Japan with the increment of areas under cedar afforestation. Although cedar pollinosis seasonally occurs, it is not disregardable in view of the public health.

In conventional therapy, for example, steroid hormone or disodium cromoglicate is administered. Such therapy is a symptomatic treatment which temporally relieves patient's symptom.

While administration of intact cedar pollen allergen responsible for cedar pollinosis has been attempted to effect hyposensitization in order to completely cure cedar pollinosis. Such hyposensitization has the drawbacks that it has a fear of eliciting anaphylaxis from the cedar pollen allergen used, and that treatment using the cedar pollen allergen should be continued for long time, i.e. about 3 years, because a small amount of the cedar pollen allergen is repeatedly administered to a cedar pollinosis patient in order to avoid such anaphylaxis.

Furthermore, cedar pollen allergen should be carefully handled because it is readily adsorbed on vessels such as glassware and metalware, and, in hyposensitization, this renders the administration of a prescribed amount of cedar pollen allergen very difficult.

Summary of the Invention

The present inventors studied modification of cedar pollen allergen in order to obtain a novel hyposensitization agent which can be used in the prevention and treatment of cedar pollinosis.

As a result, the present inventors found that a hyposensitization agent comprising a cedar pollen allergen covalently attached to a saccharide can attain the object of the present invention.

Detailed

The

of Jap

"Ura S

Asn-Pr

Asn-A

TI

amylc

gum,

and p

the r

/

non-

their

well

suc

coli

per

tiss

be

lin

25

ot

ar

h

ti

30

s

:

35

40

45

s

Detailed Description of the Invention

The cedar pollen allergen as referred to in the present invention includes those prepared from pollens of Japanese cedars (*Cryptomeria japonica*) such as "Omote Sugi (original type of Japanese cedar)" and "Ura Sugi (subspecies of Japanese cedar)", preferably, those having a partial amino acid sequence of Asp-Asn-Pro-Ile-Asp-Ser-, more particularly, Asp-Asn-Pro-Ile-Asp-Ser-X-Trp-Arg-Gly-Asp-Ser-Asn-Trp-Ala-Gln-Asn-Arg-Met-Lys- (wherein X is Ser, Cys, Thr or His) beginning at its N-terminal.

The saccharides usable in the present invention include homo- and hetero-glycans, for example, starch, amylose, dextran, polysucrose, pullulan, elsinan, curdian, gum arabic, gum tragacanth, guar gum, xanthan gum, carrageenan, pectin, cellulose, glucomannan, chitosan, and lipopolysaccharide, and their derivatives and partial hydrolysates, having an average molecular weight in the range of 500-10,000,000, preferably, in the range of 10,000-1,000,000.

A hyposensitization agent comprising a cedar pollen allergen covalently attached to a water-soluble non-ionic polysaccharide, mainly composed of repeating maltotriose units, such as pullulan, elsinan and their partial hydrolysates prevents an anaphylaxis which may be induced by intact cedar pollen allergen, as well as facilitating the preparation of a more effective hyposensitization agent of cedar pollinosis.

A hyposensitization agent comprising a cedar pollen allergen covalently attached to a hetero-glycan such as lipopolysaccharides derived from microorganisms, for example, a microorganism of the genus *E. coli*, *Salmonella* or *Serratia*, and their partial hydrolysates is favorably used as a hyposensitization agent for percutaneous and permucocutaneous administrations of cedar pollinosis because it is excellently bound to tissue such as mucous membranes.

Any procedure can be employed in the present invention as long as it forms a covalent bonding between a cedar pollen allergen and a saccharide, for example, diazo coupling-, peptide-, alkylation-, cross-linking-, disulfide-coupling-, amide-bonding-, and periodate-oxidation-methods.

In the diazo coupling method a cedar pollen allergen is allowed to react with an activated saccharide obtained by introducing an aromatic amino group, for example, p-aminobenzyl-, p-aminobezoyl-, m-aminobenzyl-, m-aminobezoyl-, m-aminoaniso-, m-aminobenzoyloxy methyl-, 3-(p-aminophenoxy)-2-hydroxy-propionyl- and 3-(p-amino-m-methyl anilino)-5-chlorotriazinyl-groups into a saccharide in conventional manner.

In the peptide method, a cedar pollen allergen is allowed to react with an activated saccharide, such as sugar carbonate and cyanogen bromide-activated saccharide, which is a derivative of a saccharide bearing a carboxyl group obtained by allowing it to react with azide, acid chloride, carbodiimide or isocyanate.

In the alkylation method, a cedar pollen allergen is allowed to react with an alkyl halide derivative of a saccharide which has been introduced with a group, for example, chloroacetyl-, bromoacetyl-, iodoacetyl- and triazinyl-halide-groups.

In the cross-linking method, a cedar pollen allergen is allowed to react with a saccharide together with a polyfunctional reagent, for example, glutaraldehyde, glyoxal, succinaldehyde, hexamethylene diisocyanate, toluene-2,4-diisocyanate, bis-azobenzidine and N,N'-ethylene-bis-maleimide.

In the amide-bonding method, a cedar pollen allergen is allowed to react with an activated saccharide which has been obtained by reacting a saccharide having an amide group with haloacylhalide, for example, bromoacetyl bromide, chlorobutyryl chloride, fluoropropionyl fluoride and iodevaleryl iodide.

The weight ratio of the cedar pollen allergen to the saccharide, both used in the covalent attachment, is usually in the range of 1:0.001-1:1,000, preferably, in the range of 1:0.01-1:100.

Any reaction conditions can be employed as long as the formation of a cedar pollen allergen-saccharide conjugate substantially does not reduce the producibilities of immunoglobulin G and M antibodies which are specific to intact cedar pollen allergen, but extremely reduces the producibility of immunoglobulin E antibody which is responsible for anaphylaxis and allergy; usually, at a temperature of about 0-100° C and a pH of about 3-12 for about 0.1-50 hours.

The cedar pollen allergen-saccharide conjugate thus obtained is usually separated and purified by conventional method, for example, filtration, washing, centrifugation, salting-out, dialysis, adsorption and desorption using ion exchange, gel filtration, ion exchange chromatography, affinity chromatography and electrophoresis into a solution and syrup, which can be dried into powder, if necessary. Thus, a hyposensitization agent of cedar pollinosis is obtained.

The hyposensitization agent can be advantageously used intact or, if necessary, in combination with a stabilizer, antiseptic agent, adjuvant and vehicle as an agent in the prevention and treatment of cedar pollinosis.

In comparison with conventional cedar pollen allergen, the hyposensitization agent prepared in this way much more enhances the producibilities of immunoglobulin G and M antibodies which are specific to intact

cedar pollen allergen, but extremely reduces the producibility of immunoglobulin E antibody which is specific to the allergen and responsible for anaphylaxis and allergy. Administration of the hyposensitization agent to a cedar pollinosis patient elicits the minimum level of immunoglobulin E antibody which is specific to the allergen.

5 In comparison with intact cedar pollen allergen, the hyposensitization agent according to the present invention has the following advantages that it is scarcely wasted because it does not adsorb on vessels such as glassware and metalware, that it is excellently stable, that it is administerable without fear of eliciting anaphylaxis and that it cuts hyposensitization period to about 1/3 to 1/200.

10 The hyposensitization agent according to the present invention is usually prepared into an injection, for example, lyophilized injection or liquid injection, and then intradermally, subcutaneously, intramuscularly or intraperitoneally administered to a cedar pollinosis patient at a dose in the range of about 0.01-100,000 ng/shot/adult about 1-2 times/week over a period of about 1-12 months to attain a prescribed hyposensitization.

15 Furthermore, the hyposensitization agent can be prepared into a form which is advantageous for percutaneous and permucocutaneous administrations, for example, troche, sublingual, tablet, ophthalmic solution, intranasal nebula, cataplasma, cream and lotion. For example, the dose and administration frequency thereof are selected such that a prescribed hyposensitization is most readily attainable.

20 In a local administration such as percutaneous and permucocutaneous administrations, the hyposensitization agent can inhibit the binding between a cedar pollen allergen and immunoglobulin E antibody which has been bound to a cedar pollinosis patient's tissue. Such action is instantly and locally occurred at area where the hyposensitization agent has been administered. Thus, the hyposensitization agent instantly relieves the patient's pain.

The hyposensitization agent can be favorably used in the prevention and treatment of cedar pollinosis elicited by "Hinoki (*Chamaecyparis obtusa*)", as well as by cedar.

25 The following experiments will explain the present invention in more detail.

Experiment I-1

30

Preparation of cedar pollen allergen

A cedar pollen collected from "Omote Sugi" grown in Chiba, Japan, was added with about 15-folds by weight of 0.125 M aqueous sodium hydrogencarbonate solution (pH 8.0). The mixture was subjected to 1 hour extraction at 4°C under gentle stirring conditions, followed by centrifugal separation. The residue was further subjected to extraction and centrifugal separation similarly as above. The resultant supernatants were pooled and salted out by the addition of ammonium sulphate to give 80% saturation, and the resultant precipitate was dialyzed and filtered. The filtrate was subjected to column chromatography using DEAE-Sephadex®, commercialized by Pharmacia LKB Biotechnology AB, Uppsala, Sweden. The unadsorbed fraction was collected, subjected to column chromatography using CM-Sephadex®, commercialized by Pharmacia LKB Biotechnology AB, Uppsala, Sweden, and eluted with phosphate buffered saline (pH 7.0). Then, the resultant solution was subjected to column chromatography using Mono S®, commercialized by Pharmacia LKB Biotechnology A3, Uppsala, Sweden, and eluted with Tris-HCl buffer (pH 7.0) to separate a solution containing a purified cedar pollen allergen exhibiting a high affinity to immunoglobulin E antibody of a cedar pollinosis patient, as well as to anti cedar pollen allergen mouse monoclonal antibody in the yield of about 0.02% against the material cedar pollen based on dry solid.

The cedar pollen allergen exhibited a molecular weight of about 50,000±5,000 on SDS-polyacrylamide gel electrophoresis, and an isoelectric point of about 8.8.

50 A partial amino acid sequence of the cedar pollen allergen was obtained by degrading it with a gas-phase protein sequencer and identifying the resultant with high-performance liquid chromatography by the method described in The Journal of Biological Chemistry, Vol. 256, pp. 7990-7997 (1981). As a result, it was found that the cedar pollen allergen had a partial amino acid sequence of Asp-Asn-Pro-Ile-Asp-Ser-X-Trp-Arg-Gly-Asp-Ser-Asn-Trp-Ala-Gln-Asn-Arg-Met-Lys- (wherein X is Ser, Cys, Thr or His).

55

Experiment I-2

Preparation

Or
added
5 5°C
solution
was
obtain
about
10 mixture
follow
stirring
CM-

15 the
ads

20 Ex

Ac

25 E

1

30

35

40

45

Preparation of cedar pollen allergen-pullulan conjugate

One hundred milliliters of 2 w/v % pullulan aqueous solution, average molecular weight of 300,000, was added with 2 ml of 1.7 w/v % cyanuric chloride in acetone. The mixture solution was allowed to stand at 5 °C or lower in ice-chilled water, adjusted to pH 7.0 by the addition of 5% sodium carbonate aqueous solution, and allowed to react for 2 hours while retaining the temperature and pH. Then, the reaction mixture was dialyzed overnight against 4 °C water while retaining the pH. Thus, an activated-pullulan solution was obtained. Thirty milliliters of the activated-pullulan solution was added with 40 ml of a solution containing about 1 mg/ml of a purified cedar pollen allergen obtained by the method in Experiment I-1. The resultant mixture was allowed to stand first at pH 7.0 and 37 °C for 5 hours while stirring, then at 5 °C overnight, followed by the addition of 6 g glycine. The resultant mixture was allowed to stand for 10 hours while stirring, dialyzed against 0.01 M acetate buffer (pH 5.0), and subjected to column chromatography using CM-Sephadex®. The unadsorbed fraction was membrane-filtered to obtain an allergen-pullulan conjugate.

The yield was about 60% against the cedar pollen allergen protein. Unlike intact cedar pollen allergen, the product is easily handleable because it is excellently stable and because it is scarcely wasted by its adsorption on glassware and metalware.

Experiment I-3Administration test on animalExperiment I-3-1Test on prophylactic activity

0.2 ml of physiological saline containing as an allergen 1 µg of an allergen-pullulan conjugate obtained by the method in Experiment I-2 was intraperitoneally administered to a group of six 10 to 12 week-old BALB/c female mice once a week over a period of 3 weeks. One-week after the intraperitoneal administration, 0.2 ml of physiological saline containing 1 µg of a cedar pollen allergen, obtained by the method in Experiment I-1, and 4 mg aluminum hydroxide as an adjuvant was administered to each mouse in the same manner as described in the above.

The amounts of immunoglobulin G, M and E antibodies which were specific to intact cedar pollen allergen were determined with a blood sample which had been collected from mice immediately before the intraperitoneal administration of the mixture of cedar pollen allergen and aluminum hydroxide, and another blood sample which had been collected from the mice 1-week after the intraperitoneal administration of the mixture.

As control, a mixture containing 1 µg of a cedar pollen allergen prepared by the method in Experiment I-1 and 40 µg of a fresh preparation of the same pullulan as used in Experiment I-2 was administered to each mouse in place of the allergen-pullulan conjugate.

The levels of immunoglobulin G and M antibodies were compared with their antibody titers determined by the technique for passive hemagglutination reaction described in Japanese Journal of Medical Science and Biology, Vol. 28, pp. 127-138 (1975), and the level of immunoglobulin E antibody was compared with its antibody titer determined by the passive cutaneous anaphylaxis reaction described in Life Science, Vol. 8, Part II, pp. 813-820 (1969). The results were as shown in Table 1.

Table 1

Period for collecting blood	Immediately before administration of mixture of cedar pollen allergen and aluminum hydroxide	One-week after administration of mixture of cedar pollen allergen and aluminum hydroxide	Note
Immunoglobulin			
Hyposensitization agent			
Cedar pollen allergen-pullulan conjugate	240	940	5
Mixture of cedar pollen allergen and pullulan	25	250	320
		Present invention	
		Control	

Annotation: Each value is an average of immunoglobulin antibody titers of immunoglobulin antibodies produced in a group of 6 mice.

ceda
hypo

Exp

Tes

me
to
aft
ce
sa

al

fi

F

25

30

35

40

As evident from the results in Table 1, unlike the mixture of cedar pollen allergen and pullulan, the cedar pollen allergen-pullulan conjugate according to the present invention can be favorably used as a hyposensitization agent in the prevention of cedar pollinosis.

Experiment I-3-2

Test on therapeutic activity

0.2 ml of physiological saline containing a mixture of 1 μ g of a cedar pollen allergen obtained by the method in Experiment I-1 and 4 mg aluminum hydroxide as an adjuvant was intraperitoneally administered to a group of six 10 to 12 week-old BALB/c female mice once a week over a period of 3 weeks. Two-weeks after the intraperitoneal administration, 0.2 ml of physiological saline containing as an allergen 1 μ g of a cedar pollen allergen-pullulan conjugate obtained by the method in Experiment I-2 was administered in the same manner as described in the above to each mouse 3-times a week over a period of 3 weeks.

Formation of immunoglobulin E antibody was boosted by administering a mixture of cedar pollen allergen and aluminum hydroxide to the mice.

The levels of immunoglobulin G, M and E antibodies were determined with a blood sample collected from mice just before and 1-week after the final intraperitoneal administration of the cedar pollen allergen-pullulan conjugate, and another blood sample which had been collected from mice 1-week after the induction of immunoglobulin E antibody by the intraperitoneal administration of the mixture of cedar pollen allergen and aluminum hydroxide.

As control, a mixture of cedar pollen allergen and pullulan was used similarly as in Experiment I-3-1 in place of the cedar pollen allergen-pullulan conjugate. The results were as shown in Table 2.

Table 2

Period collecting blood	Immediately before ad- ministration of cedar pollen allergen-pullulan conjugate	One-week after ad- ministration of cedar pollen allergen- pullulan conjugate	One-week after production of immunoglobulin E antibody by booster shot	Note			
Immunoglobulin							
Hyposensitization agent							
Cedar pollen allergen- pullulan conjugate	350	160	2,300	30	5,840	30	Present invention
Mixture of cedar pollen allergen and pullulan	350	160	480	320	2,870	1,280	Control

Annotation: Each value is an average of immunoglobulin antibody titers of immunoglobulin antibodies produced in a group of 6 mice.

5
10
15
20
25
30
35
40
45
50
55

pollen
sitiz
hyc
cer
the
all
in
pc
E
F
E
30
35

As evident from the results in Table 2, unlike a mixture of cedar pollen allergen and pullulan, the cedar pollen allergen-pullulan conjugate according to the present invention can be favorably used as a hyposensitization agent in the treatment of cedar pollinosis.

Mice which had been previously primed with a mixture of cedar pollen allergen and aluminum hydroxide to produce immunoglobulin E antibody were sprayed with a physiological saline containing the cedar pollen allergen-pullulan conjugate into their mouths and nasal cavities. One hour after the spraying, the mice were resprayed with intact cedar pollen allergen into their mouths and nasal cavities, and the allergic reaction which would be elicited in the mice was not observed.

As described hereinbefore, a cedar pollen allergen-saccharide conjugate according to the present invention can be favorably used as a hyposensitization agent in the prevention and treatment of cedar pollinosis because the conjugate effects a high hyposensitivity without fear of eliciting anaphylaxis.

Experiment II-1

Preparation of cedar pollen allergen and lipopolysaccharide conjugate

One milliliter of 10 mM calcium phosphate solution containing 10 mg of lipopolysaccharide derived from a microorganism of the species *E. coli* was added with 60 ml of 100 mM sodium periodate, and the mixture was allowed to react at ambient temperature for 20 minutes to cleave the specified bonding of the saccharide chain in the lipopolysaccharide. Then, the resultant mixture was dialyzed overnight against 1 M glycine-HCl buffer (pH 4.4) at 4°C to remove an excess amount of sodium periodate. The resultant solution was adjusted to about pH 9.5 by the addition of 0.1 M sodium hydrogencarbonate buffer to prepare a lipopolysaccharide solution.

Ten milligrams of a cedar pollen allergen prepared by the method in Experiment I-1 was dissolved in 1 ml of phosphate buffer (pH 9.5), and the mixture was added with the lipopolysaccharide solution, followed by the formation of Schiff base of the cedar pollen allergen and the lipopolysaccharide.

The reaction mixture was added with sodium borohydride to complete coupling reaction. The resultant solution was subjected to column chromatography using Sephadex® G-100, a product of Pharmacia LKB Biotechnology AB, Uppsala, Sweden, and the fraction containing a cedar pollen allergen-lipopolysaccharide conjugate was collected. The fraction was membrane-filtered to obtain the cedar pollen allergen-lipopolysaccharide conjugate.

The yield of the product was about 40% against the cedar pollen allergen protein.

Unlike intact cedar pollen allergen, the product is easily handleable and scarcely wasted because it does not adsorb on vessels such as glassware.

Experiment II-2

Administration test on animal

Test on therapeutic activity

One milliliter of physiological saline containing as an allergen 10 µg of a cedar pollen allergen-lipopolysaccharide conjugate prepared by the method in Experiment II-1 was orally administered to a group of six 10 to 12 week-old BALB/c female mice 3-times a week over a period of 3 weeks. One-week after the oral administration, blood samples were collected from the mice, and the amounts of immunoglobulin A, G and E antibodies which were specific to intact cedar pollen allergen were determined with the blood samples.

As control, a mixture of intact cedar pollen allergen and lipopolysaccharide was orally administered to mice similarly as above in place of the cedar pollen allergen-lipopolysaccharide conjugate. The levels of

immunoglobulin A and G antibodies were compared with their antibody titers determined by the enzyme immunoassay method described in Journal of Immunological Methods, Vol. 6, pp. 355-362 (1975), while the level of immunoglobulin E antibody was compared with its antibody titer determined by the passive cutaneous anaphylaxis reaction described in Experiment 1-3-1. The results were as shown in Table 3.

Table 3

Immunoglobulin	Hypersensitization agent			Note
	A	G	E	
Cedar pollen allergen-lipopolysaccharide conjugate	256	64	0	Present invention
Mixture of cedar pollen allergen and lipopolysaccharide	4	16	16	Control

Annotation: Each value is an average of immunoglobulin antibody titers of immunoglobulin antibodies produced in a group of 6 mice.

As evident from the results in Table 3, unlike the mixture of cedar pollen allergen and lipopolysaccharide, the cedar pollen allergen-lipopolysaccharide conjugate according to the present invention can be favorably used as a hyposensitization agent in the prevention and treatment of cedar pollinosis.

Mice which had been previously primed with a mixture of cedar pollen allergen and aluminum hydroxide to produce immunoglobulin E antibody were sprayed with a physiological saline containing the cedar pollen allergen-lipopolysaccharide conjugate into their mouths and nasal cavities. Thirty minutes after the spraying, the mice were resprayed with intact cedar pollen allergen into their mouths and nasal cavities, and the allergic reaction which would be elicited in the mice was not observed.

A cedar pollen allergen-hetero glycan conjugate such as a cedar pollen allergen-lipopolysaccharide conjugate is superior in absorbability into mucous membranes to a cedar pollen allergen-homo glycan conjugate such as a cedar pollen allergen-pullulan conjugate because the cedar pollen allergen-lipopolysaccharide conjugate is adsorbed on the mucous membranes for a relatively long period of time. Because of the above reason, such cedar pollen allergen-hetero glycan conjugate is favorably used as a hyposensitization agent for percutaneous and permucocutaneous administrations such as oral and intranasal administrations.

Several embodiments according to the present invention will hereinafter be explained.

Example 1

Example 1(1)

Preparation of cedar pollen allergen

A cedar pollen collected from "Ura Sugi" grown in Akita, Japan, was prepared into a purified cedar pollen allergen in solution by the method in Experiment I-1 in the yield of about 0.015% against the weight of the material cedar pollen based on dry solid. The purified cedar pollen allergen had a molecular weight of about 50,000 \pm 5,000 on SDS-polyacrylamide gel electrophoresis, and an isoelectric point of about 8.8.

After detecting a partial amino acid sequence of the cedar pollen allergen in accordance with the method in Experiment I-1, the partial amino acid sequence consisting of 20-amino acid residues beginning at its N-terminal was identical with that of the allergen prepared from "Omote Sugi" described in Experiment I-1.

Example 1(2)

Hyposensitization agent

Five grams of pullulan having an average molecular weight of about 140,000 was dissolved in 400 ml of water, and the resultant solution was adjusted to pH 10.7 by the addition of 1 N sodium hydroxide. The solution was then allowed to react with 3 g cyanogen bromide for 1 hour while gradually adding it to the solution and retaining the pH. The reaction mixture was adjusted to pH 5.0 by the addition of 1 N hydrochloric acid, and dialyzed against cold water while retaining the pH. Thus, an activated-pullulan solution was obtained.

To the activated-pullulan solution was added 200 ml of a cedar pollen allergen solution prepared by the method in Example 1(1), and the mixture was allowed to react at ambient temperature for 24 hours. After completion of the reaction, the reaction mixture was added with acetone (1:3 by volume) to obtain a precipitate which was then collected, dissolved in 0.01 M acetate buffer (pH 5.0), and centrifugally separated to remove the insoluble residue. The remaining supernatant was subjected to column chromatography using CM-Sephadex®, and the unadsorbed fraction was membrane-filtered. The filtrate was bottled into ampules to obtain a liquid hyposensitization agent containing a cedar pollen allergen-pullulan conjugate.

The yield was about 70% against the cedar pollen allergen protein. The product can be favorably used

in the prevention and treatment of cedar pollinosis because the product effects a high hyposensitivity without fear of eliciting anaphylaxis.

Unlike intact cedar pollen allergen, the product is easily handleable because it is excellently stable and because it is scarcely wasted by its adsorption on glassware and metalware.

Example 2

Fifty-two grams of a pullulan partial hydrolysate, average molecular weight of about 10,000, was dissolved in 110 ml of dimethylformamide while heating. The mixture was cooled to ambient temperature, and added with 10 ml of pyridine. The resultant mixture was added with 1.0 g 4-nitrobenzoyl chloride while stirring, and allowed to stand at ambient temperature for 17 hours. The reaction mixture was added with 2-volumes of n-propyl alcohol to obtain a precipitate which was then collected and dissolved in dimethylformamide. The above precipitation was repeated 3-times, and the resultant precipitates were pooled and dissolved in 100 ml of 5 w/v % sodium hydrosulfite aqueous solution. The mixture was allowed to stand at 80° C for 30 minutes, decolorized with activated charcoal and precipitated with 2-volumes of n-propyl alcohol. The precipitate was dissolved in water, and the resultant solution was dialyzed overnight against water, cooled to 2° C or lower, and added with hydrochloric acid to give a final concentration of about 0.1 N while stirring. Then, the resultant mixture was added with sodium nitrite to give a concentration of about 1 w/v %, allowed to react for 30 minutes, and dialyzed against distilled water at 2° C or lower for 2 hours to obtain an activated pullulan partial hydrolysate.

To the activated pullulan partial hydrolysate was added 20 ml of a solution containing a cedar pollen allergen from "Omote Sugi" obtained by the method in Experiment I-1, and the mixture was adjusted to pH 8.5 by the addition of a sodium carbonate aqueous solution. Then, the mixture was allowed to effect coupling reaction at 4° C for 2 hours while stirring, purified similarly as in Example 1, and bottled into ampules to obtain a liquid hyposensitization agent containing a cedar pollen allergen-pullulan partial hydrolysate conjugate.

The yield was about 60% against the cedar pollen allergen protein.

Similarly as the product in Example 1, the liquid hyposensitization agent is favorably used in the prevention and treatment of cedar pollinosis and is easily handleable because it is excellently stable and because it is scarcely wasted by its adsorption on glassware and metalware.

Example 3

Ten grams of elsinan, average molecular weight of about 200,000, was dissolved in 200 ml of distilled water while heating. The resultant was cooled to ambient temperature and added with 5 g hexamethylenediamine. The mixture solution was adjusted to pH 11.0 by the addition of 1 N sodium hydroxide solution, allowed to stand at 20° C or lower in ice-chilled water, and added with 5 g cyanogen bromide while retaining the temperature and pH. The resultant mixture was allowed to react for 30 minutes while stirring and retaining the temperature and pH. The reaction mixture was dialyzed against 4° C distilled water for 1 hour to obtain an activated-elsinan solution.

The activated-elsinan solution was added with 2 ml of 25 w/v % glutaraldehyde and 60 ml of a solution containing a cedar pollen allergen from "Omote Sugi" obtained by the method in Experiment I-1, and the mixture was added with 10 ml of 1 M acetate buffer (pH 5.0) to effect coupling reaction at 4° C for 24 hours while stirring. The reaction mixture was added with glycine to give a concentration of 1 M, and the resultant was allowed to stand at ambient temperature for 24 hours, and then subjected to centrifugal separation. The supernatant was subjected to gel filtration, and a fraction containing a cedar pollen allergen-elsinan conjugate was collected. The fraction was concentrated and membrane-filtered. The filtrate was bottled, lyophilized and sealed to obtain a solid hyposensitization agent containing the cedar pollen allergen-elsinan conjugate.

The yield was about 50% against the cedar pollen allergen protein.

Similarly as the product in Example 1, the product is easily handleable and usable in the prevention and treatment of cedar pollinosis.

Example 4

Two hundred milliliters of 1 w/v % carboxymethyl cellulose solution, average molecular weight of about 20,000, was added with 2 g 1-ethyl-3-(3-dimethylaminopropyl)-carbodiimide-methiodide, and the mixture solution was adjusted to pH 4.0 and allowed to react at ambient temperature for 2 hours while stirring and retaining the pH. The reaction mixture was dialyzed overnight against distilled water to obtain an activated-carboxymethyl-cellulose solution.

The activated-carboxymethyl-cellulose solution was added with 50 ml of a solution containing a cedar pollen allergen from "Ura Sugi" obtained by the method in Example 1(1), and the activated carboxymethyl cellulose and the cedar pollen allergen were allowed to effect coupling reaction at ambient temperature overnight while stirring and retaining pH at 4.5. Then, the resultant conjugate was purified similarly as the method in Example 3, and bottled into ampules to obtain a liquid hyposensitization agent containing the cedar pollen allergen-carboxymethyl cellulose conjugate.

The yield was about 30% against the cedar pollen allergen protein.

Although the product is slightly lower in producibilities of immunoglobulin G and M antibodies which are specific to intact cedar pollen allergen than a cedar pollen allergen-pullulan conjugate and a cedar pollen allergen-elsinan conjugate, the product is easily handleable because it does not produce immunoglobulin E antibody which is specific to the allergen, and because it is usable as a hyposensitization agent in the prevention and treatment of cedar pollinosis.

Example 5

One hundred milligrams of a lipopolysaccharide derived from a microorganism of the genus Salmonella was added with 25 ml of 50% saturated sodium acetate at about 4 °C, and the mixture was adjusted to pH 9.0 by the addition of 0.5 N sodium hydroxide. To the resultant mixture was added dropwise a mixture solution (about pH 8.5) of 20 μ l bromoacetyl bromide and 1 ml anhydrous dioxane. The obtained mixture was adjusted to about pH 4.5 by the addition of 6 N hydrochloric acid, and dialyzed against 4 °C water for 5 days to obtain an activated-lipopolysaccharide solution. The activated-lipopolysaccharide solution was added with 40 ml of a cedar pollen allergen solution prepared by the method in Example 1(1), and the mixture was allowed to react at 25 °C for 2 days while stirring and retaining its pH at 4.5. The reaction mixture was purified similarly as in Example 3, and bottled into ampules to obtain a liquid hyposensitization agent containing a cedar pollen allergen-lipopolysaccharide conjugate.

The yield of the product was about 35% against the cedar pollen allergen protein.

The product is easily absorbed into mucous membranes because it is excellently bound to them and locally adsorbed on them for a relatively long period of time. The product is an excellent hyposensitization agent of cedar pollinosis for percutaneous and permucocutaneous administrations such as oral and intranasal administrations.

Effect of the invention

As evident from the above, a hyposensitization agent comprising a cedar pollen allergen covalently attached to a saccharide is administrable to a cedar pollinosis patient without fear of eliciting anaphylaxis, and cuts hyposensitization period to about 1/3 to 1/200 because the hyposensitization agent extremely accelerates the production of immunoglobulin G and M antibodies which are specific to intact cedar pollen allergen, but extremely reduces the production of immunoglobulin E antibody which is specific to the allergen.

The hyposensitization agent according to the present invention is favorably used as an agent for local administrations such as percutaneous and permucocutaneous administrations because the hyposensitization agent inhibits the antigen-antibody reaction between immunoglobulin E antibody which has been bound to a cedar pollinosis patient's tissue and intact cedar pollen allergen, and instantly relieves the patient's pain.

Furthermore, the hyposensitization agent according to the present invention has a great significance in the field from the viewpoint of that it is scarcely wasted because it does not adsorb on vessels such as glassware and metalware, that it is excellently stable, and that it is easily handleable as compared with

intact cedar pollen allergen.

While there has been described what is at present considered to be the preferred embodiments of the invention, it will be understood that various modifications may be made therein, and it is intended to cover in the appended claims all such modifications as fall within the true spirit and scope of the invention.

5

Claims

1. A hyposensitization agent, which comprises a cedar pollen allergen covalently attached to a
10 saccharide.

2. A hyposensitization agent as claimed in claim 1, wherein said cedar pollen allergen comprises a partial amino acid sequence of Asp-Asn-Pro-Ile-Asp-Ser- beginning at the N-terminal of said cedar pollen allergen.

3. A hyposensitization agent as claimed in claim 1 or 2, wherein said cedar pollen allergen comprises a
15 partial amino acid sequence of Asp-Asn-Pro-Ile-Asp-Ser-X-Trp-Arg-Gly-Asp-Ser-Asn-Trp-Ala-Gln-Asn-Arg-Met-Lys- (wherein X is Ser, Cys, Thr or His) beginning at the N-terminal of said cedar pollen allergen.

4. A hyposensitization agent as claimed in claim 1, 2 or 3, wherein said cedar pollen allergen can be prepared from the pollen of Cryptomeria Japonica.

5. A hyposensitization agent as claimed in any one of the preceding claims, wherein said saccharide is
20 a polysaccharide mainly composed of repeating maltotriose units.

6. A hyposensitization agent as claimed in any one of claims 1 to 5, wherein said saccharide is a lipopolysaccharide.

7. A hyposensitization agent as claimed in any one of the preceding claims, wherein said agent is for
25 treatment of cedar pollinosis.

30

35

40

45

50

55



Cate:

A



European Patent
Office

EUROPEAN SEARCH REPORT

Application Number

EP 88 30 8370

DOCUMENTS CONSIDERED TO BE RELEVANT			
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int. Cl.4)
A	CHEMICAL ABSTRACTS, vol. 90, no. 5, 29th January 1979, page 374, abstract no. 37533k, Columbus, Ohio, US; G.N. GROSS et al.: "Isolation and partial characterization of the allergen in mountain cedar pollen" & SCAND. J. IMMUNOL. 1978, 8(5), 437-41 * Abstract * ---	1-7	A 61 K 39/36 C 07 K 15/10 A 61 K 47/00
A	PATENT ABSTRACTS OF JAPAN, vol. 9, no. 171 (C-291)[1894], 16th July 1985; & JP-A-60 42 335 (TORII YAKUHIN K.K.) 06-03-1985 * Abstract * -----	1-7	
			TECHNICAL FIELDS SEARCHED (Int. Cl.4)
			A 61 K
The present search report has been drawn up for all claims			
Place of search THE HAGUE		Date of completion of the search 02-11-1988	Examiner FERNANDEZ Y BRANAS F.J.
CATEGORY OF CITED DOCUMENTS			
X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document			

FORM 1503 03.82 (10/80)

11. An arrangement according to claim 9 or claim 10, in which the viewing screen is on an incline and a mask in the optical path trims the image to fit the inclined viewing screen.

12. An arrangement according to any one of claims 1 to 8, comprising a microscope arrangement of which an eye-piece is situated in the console at the viewing position. 5

13. An arrangement according to claim 11, the microscope arrangement comprising a reflected light arrangement. 10

14. An arrangement according to claim 11 or claim 12, the microscope arrangement comprising a transmitted light arrangement.

15. An arrangement according to any one of claims 1 to 13, adapted for observation of crystal melting, and formation comprising a warm air jet arrangement operable from the console to direct warm air at a crystal specimen in the specimen position to melt the same. 15 20

16. An arrangement according to any one of claims 1 to 14, comprising a live specimen chamber comprising a microscope slide with a specimen region surrounded by a wall of material which is oxygen - but not water - permeable and topped with a cover slip. 25

17. An arrangement according to claim 15, in which the wall does not completely surround the specimen area, there being a removable closure.

18. An arrangement according to any one of claims 1 to 16, comprising a live specimen chamber with food, water and air supply means for the chamber. 30

35

40

45

50

55